

# Biotechnological Applications of Elementary Modes Analysis

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# Outline

Outline

● **Outline**

Pathway Design for  
Improving Yield

*Acetobacterium woodii*  
and the  
Woods-Ljungdahl  
Pathway

Channelling Metabolism  
into Desired Routes

Conclusion

Some examples of applications of Elementary Modes Analysis:

- Pathway redesign for improving yield
- Modes of operation of the Woods–Ljungdhal pathway
- Channelling metabolism into desired routes

## Outline

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### Pathway Design for Improving Yield

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- Biodegradable Plastics
- Polyhydroxybutyrate Synthesis in Yeast
- Optimal yields of PHB synthesis

*Acetobacterium woodii*  
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Conclusion

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# Pathway Design for Improving Yield

# Biodegradable Plastics

## Outline

### Pathway Design for Improving Yield

#### ● Biodegradable Plastics

#### ● Polyhydroxybutyrate Synthesis in Yeast

#### ● Optimal yields of PHB synthesis

### *Acetobacterium woodii* and the Woods-Ljungdahl Pathway

### Channelling Metabolism into Desired Routes

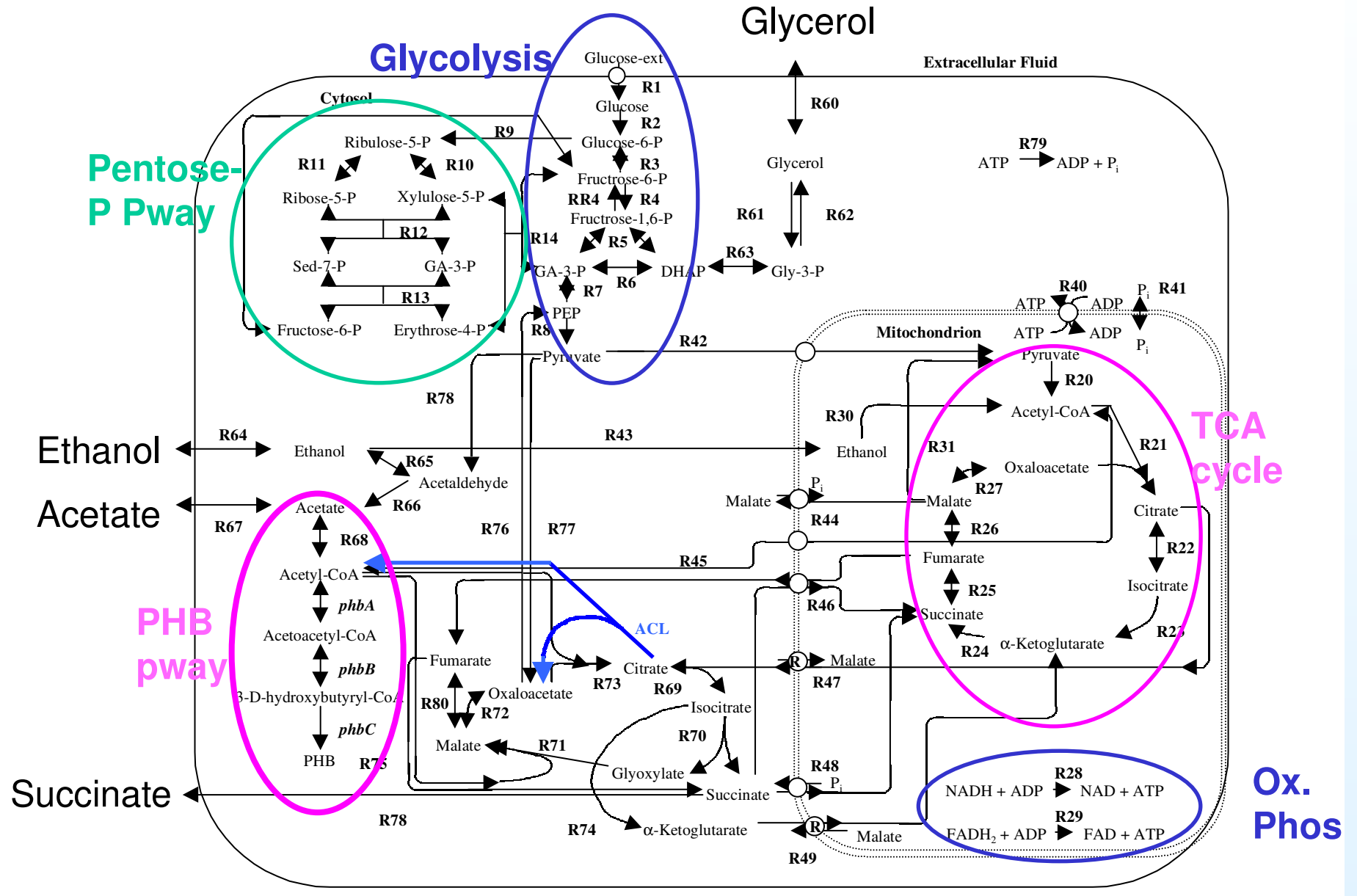
## Conclusion

## Polyhydroxybutyrate (PHA) or polyhydroxyalkanoates PHB



PHB is produced naturally as a storage compound by some bacteria, and notably by the H<sub>2</sub>-utilising, CO<sub>2</sub>-fixing *Cupriavidus necator* (aka *Ralstonia eutropha*).

# Polyhydroxybutyrate Synthesis in Yeast



# Optimal yields of PHB synthesis

Based on highest-yielding elementary modes of the network:

## Wild-type yeast + PHB pathway

1.  $2 \text{ Acetate} + \text{EtOH} \rightarrow \text{PHB} + 2 \text{ CO}_2$       0.67
2.  $65 \text{ Ac.} + 31 \text{ EtOH} \rightarrow 30 \text{ PHB} + 72 \text{ CO}_2$       0.63

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(Number following each mode is the fractional carbon conversion.)

Carlson et al, *Biotechnol. Bioeng.* **79**, 121–134, 2002.

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# Optimal yields of PHB synthesis

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## Wild-type yeast + ATP-citrate lyase + PHB pathway

3.  $12 \text{ EtOH} \rightarrow 5 \text{ PHB} + 4 \text{ CO}_2$       0.83
4.  $77 \text{ EtOH} + 31 \text{ Glycerol} \rightarrow$   
 $48 \text{ PHB} + 4 \text{ Ac.} + 47 \text{ CO}_2$       0.78

(Number following each mode is the fractional carbon conversion.)

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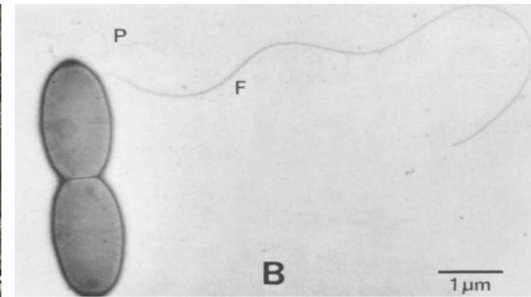
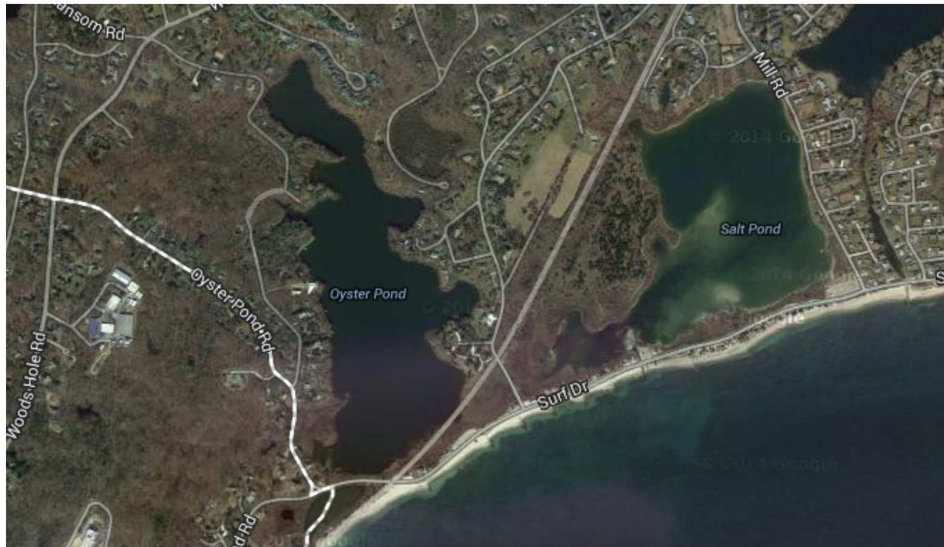
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# ***Acetobacterium woodii* and the Woods-Ljungdahl Pathway**



# *Acetobacterium Woodii*



Isolated from high salinity lake ~ 32ppt

Non-pathogenic, highly motile, non spore forming and gram positive.

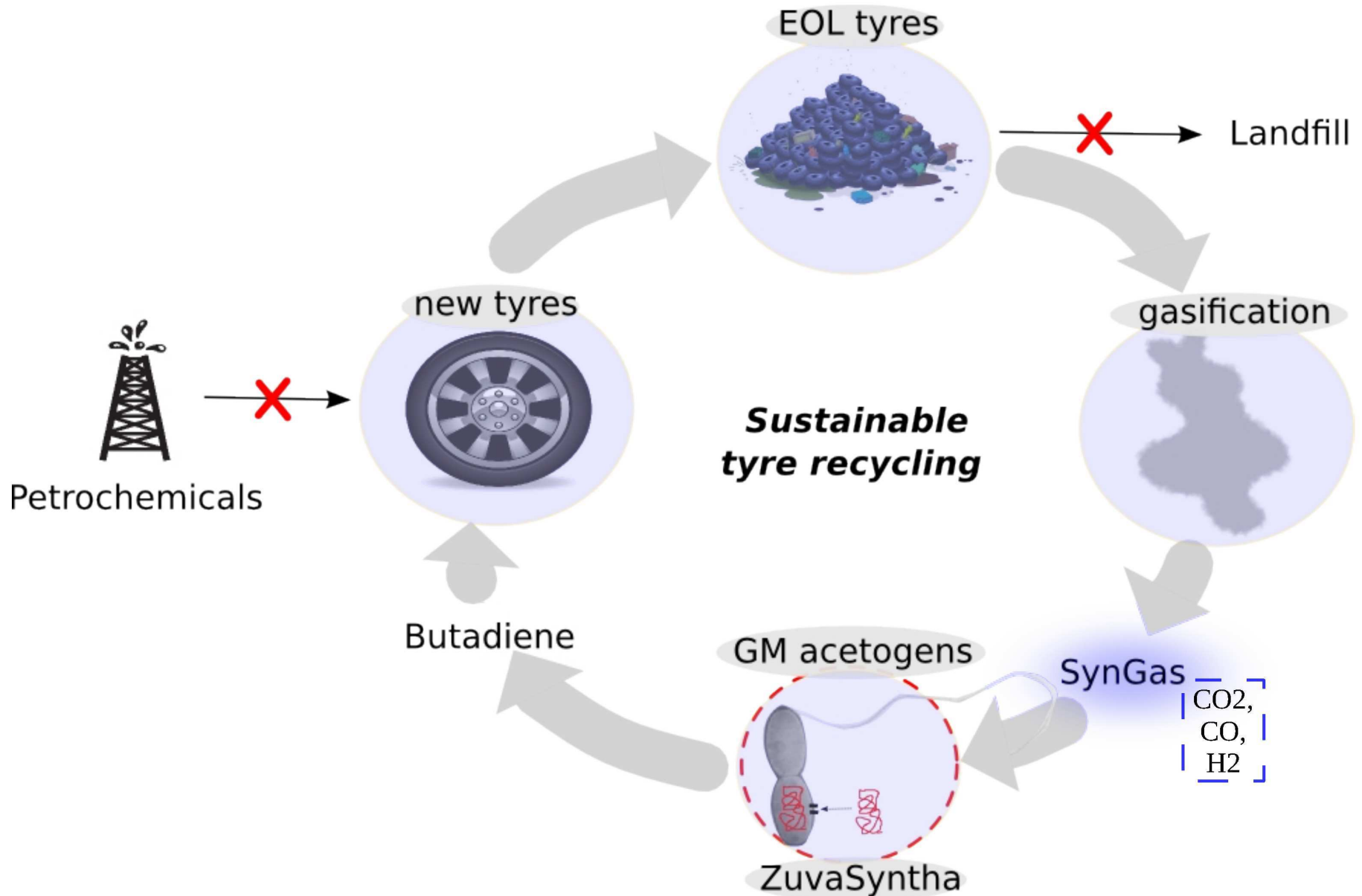
Within group of acetogens – 22 genera in soils, sediments, intestinal tracts.

Acetogens use the acetyl-CoA or Wood-Ljungdahl Pathway to grow autotrophically on inorganic substrates (H<sub>2</sub>-CO<sub>2</sub>, CO) but also metabolise organic substrates – hexoses, pentoses, alcohols, methyl groups.

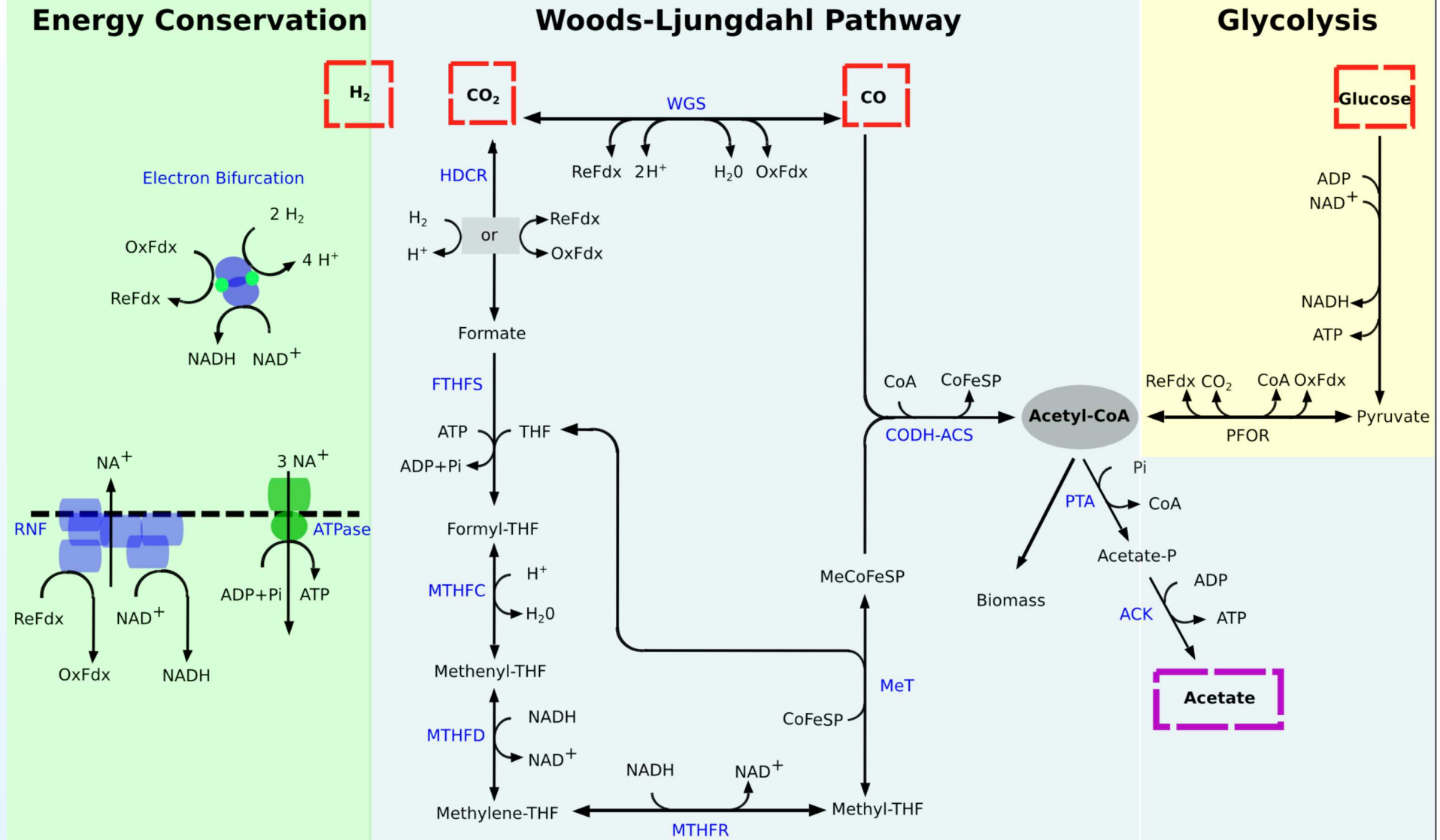
Model acetogens – *Moorella thermoacetica*, *Acetobacterium Woodii* and *Clostridium Ljungdahlii*

Currently categorised as a RNF Na-dependant acetogen  
Genome - 4,044,777 bp, 3,473 proteins

# Industrial Application



# Catabolism Overview



# The WLP Reaction Scheme

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### Pathway Design for Improving Yield

### *Acetobacterium woodii* and the Woods-Ljungdahl Pathway

- Characteristics

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- **The WLP Reaction Scheme**

- Key to reaction scheme

- Elementary modes of the WLP

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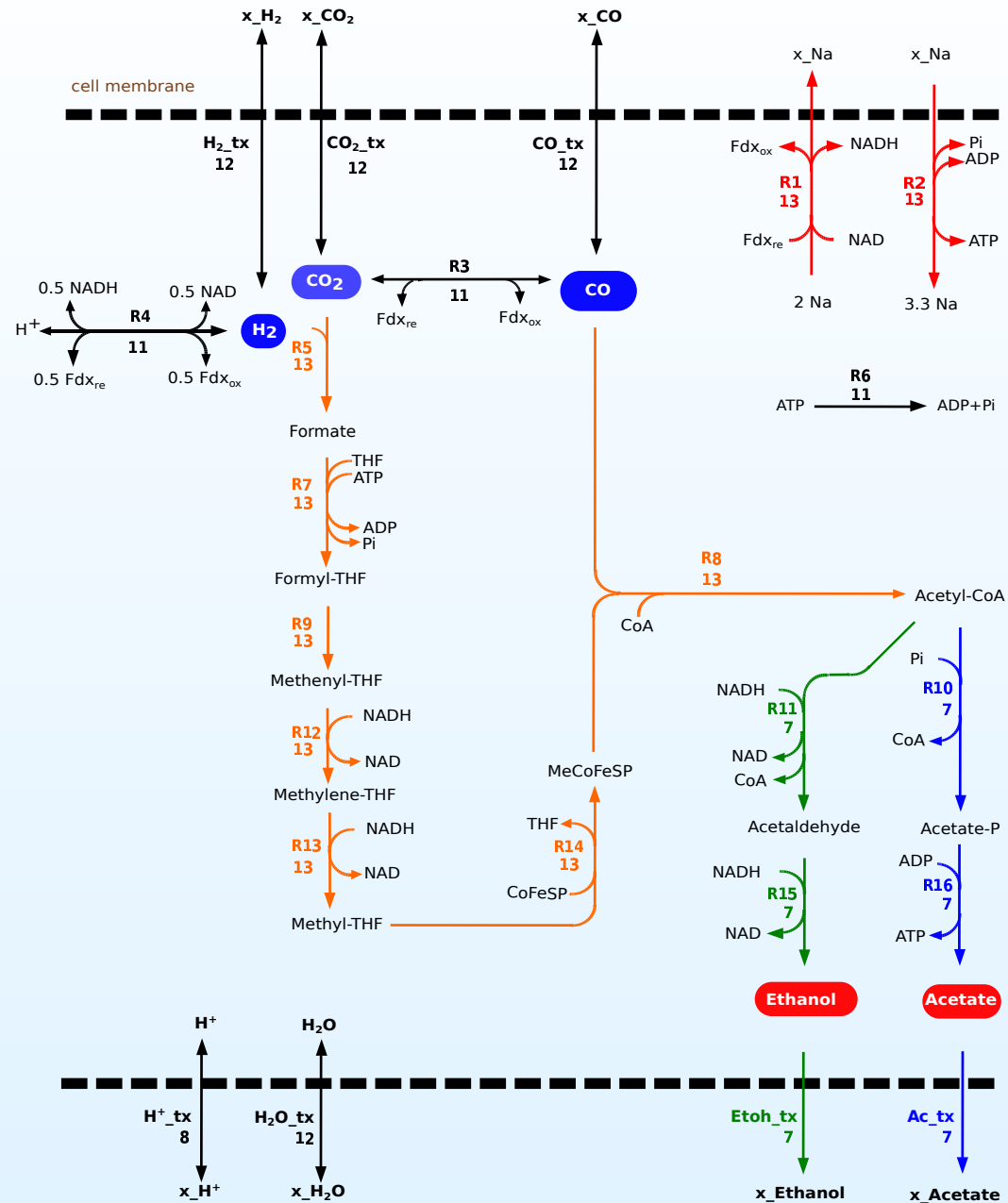
- Mode 10:  $\text{CO} + \text{H}_2$

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### Channelling Metabolism into Desired Routes

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## Key to reaction scheme

Metabolites with blue backgrounds have exchange transporters and those with red backgrounds have export transporters.

Dashed lines represent the cell membrane and so metabolites inside these are internal metabolites. The colour of non-black arrows represents membership to one of the four reaction subsets.

Numbers under reaction labels indicate the number of elementary modes that reaction is involved in.

**R1**:Rnf complex, **R2**:ATP synthase, **R3**:carbon monoxide dehydrogenase (CODH), **R4**:bifurcating hydrogenase (EBHyd), **R5**:hydrogen-dependent formate dehydrogenase (HDFD), **R6**:ATP hydrolase, **R7**:formate-THF ligase, **R8**:acetyl-CoA synthase, **R9**:methenyl-THF cyclohydrolase, **R10**:phosphate acetyltransferase, **R11**:acetaldehyde dehydrogenase, **R12**:methylene-THF dehydrogenase, **R13**:methyl-THF reductase, **R14**:methyl-THF:corrinoide/iron-sulfur methyltransferase, **R15**:alcohol dehydrogenase, **R16**:acetate kinase, **Ac\_tx**:acetate transporter, **EtoH\_tx**:ethanol transporter.

## Elementary modes of the WLP

Mode	Stoichiometry	Reactions	$Y_{ATP}^C$
1	$2 \text{ CO}_2, 4 \text{ H}_2 \rightarrow \text{Ac}, 2 \text{ H}_2\text{O}, \text{ H}^+$	19	0.15
2	$26 \text{ CO}_2, 72 \text{ H}_2 \rightarrow 3 \text{ Ac}, 10 \text{ EtOH}, 36 \text{ H}_2\text{O}, 3 \text{ H}^+$	19	0.00
3	$3 \text{ CO}_2, 5 \text{ H}_2 \rightarrow \text{Ac}, \text{ CO}, 3 \text{ H}_2\text{O}, \text{ H}^+$	15	0.00
4	$4 \text{ CO}, 2 \text{ H}_2\text{O} \rightarrow \text{Ac}, 2 \text{ CO}_2, \text{ H}^+$	18	0.38
5	$\text{CO}, \text{ H}_2\text{O} \rightarrow \text{H}_2, \text{ CO}_2$	9	0.30
6	$6 \text{ CO}, 3 \text{ H}_2\text{O} \rightarrow \text{EtOH}, 4 \text{ CO}_2$	18	0.29
7	$\text{CO}_2, \text{ CO}, 3 \text{ H}_2 \rightarrow \text{Ac}, \text{ H}_2\text{O}, \text{ H}^+$	19	0.30
8	$\text{CO}_2, \text{ CO}, 5 \text{ H}_2 \rightarrow \text{EtOH}, 2 \text{ H}_2\text{O}$	18	0.11
9	$17 \text{ CO}_2, 3 \text{ CO}, 57 \text{ H}_2 \rightarrow 10 \text{ EtOH}, 27 \text{ H}_2\text{O}$	18	0.00
10	$2 \text{ CO}, 2 \text{ H}_2 \rightarrow \text{Ac}, \text{ H}^+$	18	0.45
11	$3 \text{ CO}, \text{ H}_2, \text{ H}_2\text{O} \rightarrow \text{Ac}, \text{ CO}_2, \text{ H}^+$	19	0.40
12	$5 \text{ CO}, \text{ H}_2, 2 \text{ H}_2\text{O} \rightarrow \text{EtOH}, 3 \text{ CO}_2$	18	0.29
13	$3 \text{ CO}, 3 \text{ H}_2 \rightarrow \text{EtOH}, \text{ CO}_2$	18	0.27
14	$2 \text{ CO}, 4 \text{ H}_2 \rightarrow \text{EtOH}, \text{ H}_2\text{O}$	18	0.26

# Mode 1: CO<sub>2</sub> + H<sub>2</sub>

## Outline

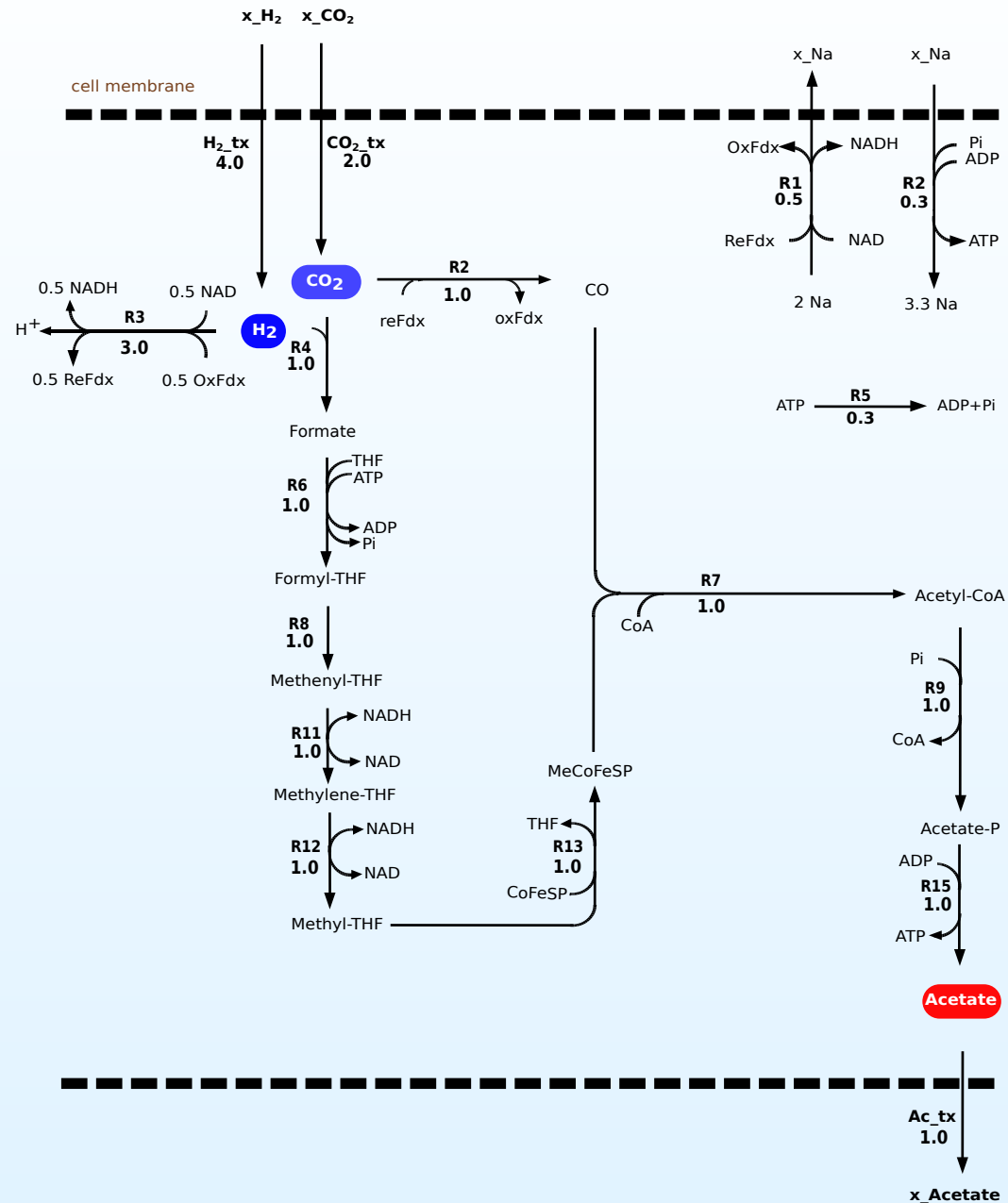
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#### *Acetobacterium woodii* and the Woods-Ljungdahl Pathway

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### Channelling Metabolism into Desired Routes

## Conclusion





# Mode 4: CO + H<sub>2</sub>O

## Outline

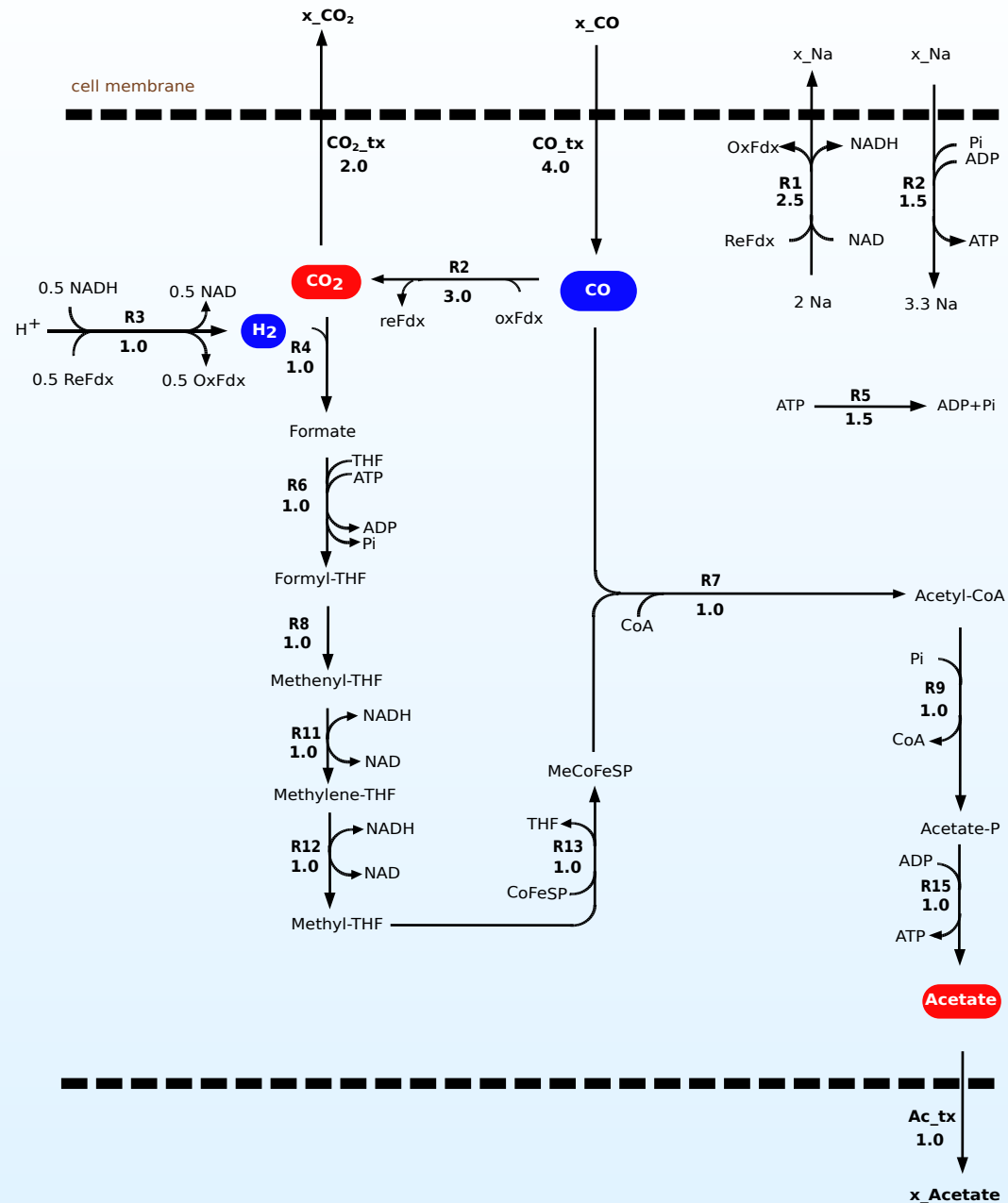
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# Mode 7: CO<sub>2</sub> + CO + H<sub>2</sub>

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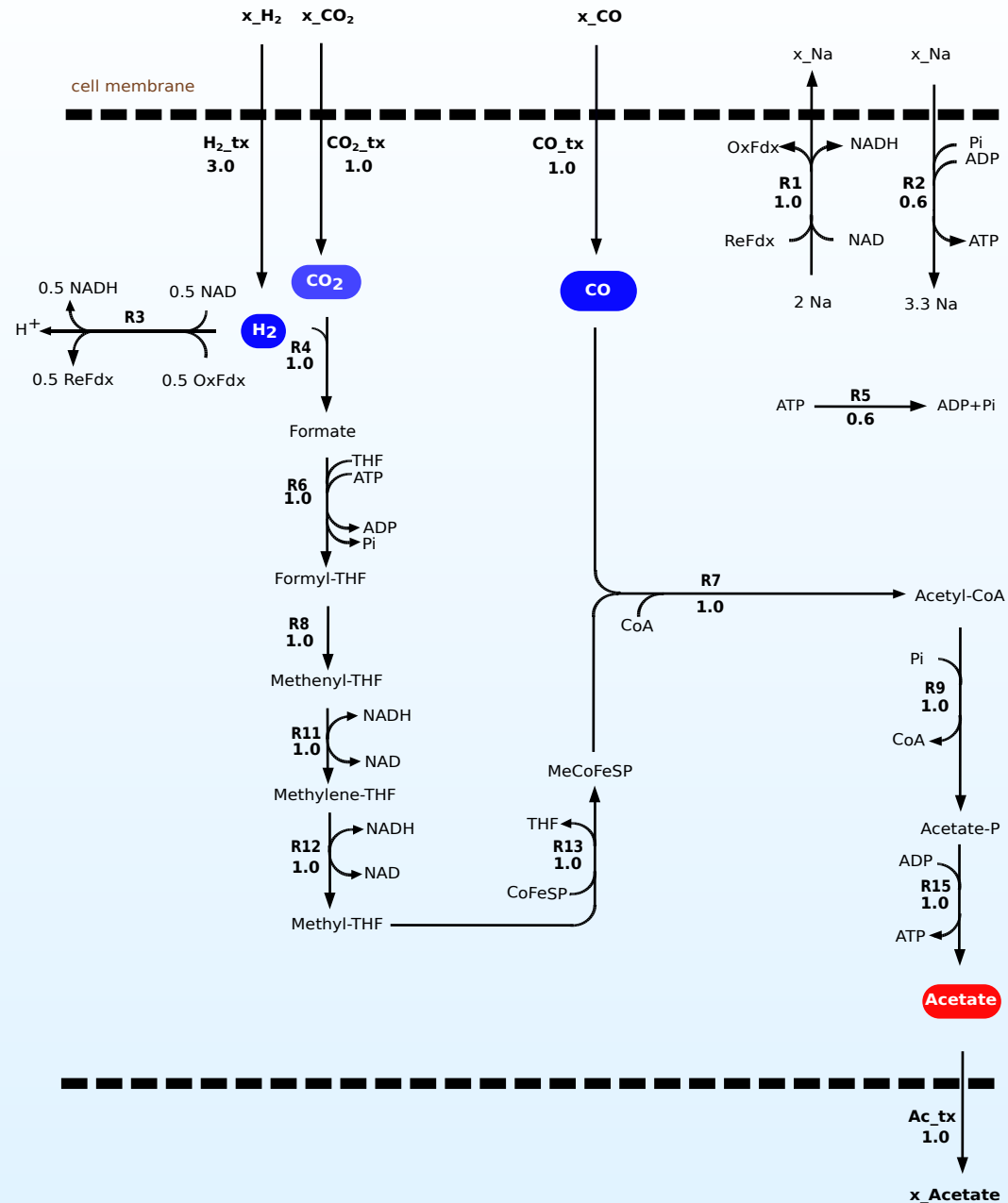
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# Mode 10: CO + H<sub>2</sub>

## Outline

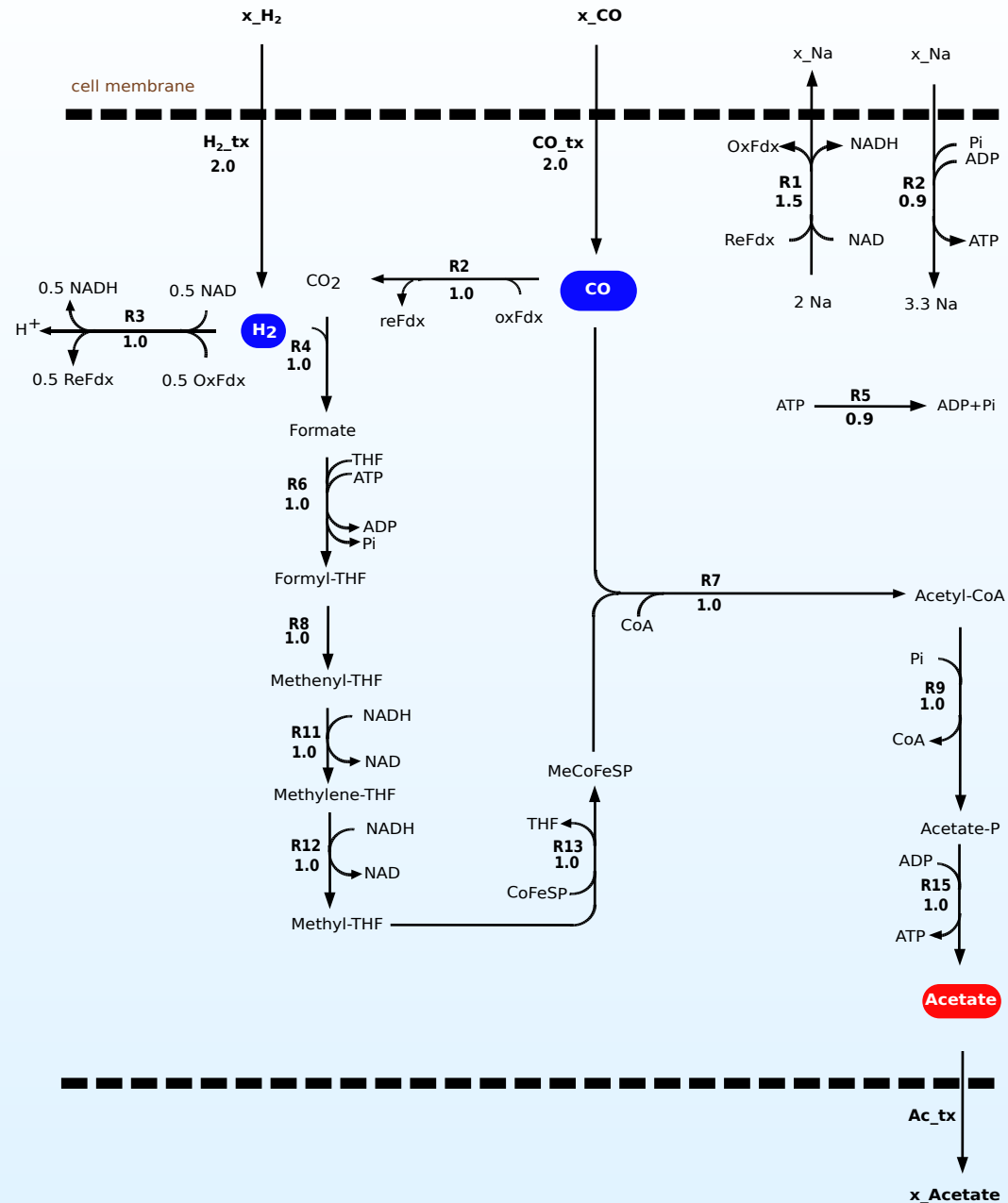
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## An aside — *C. autoethanogenum*

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### Conclusion

- *Clostridium ethanogenum* uses the WL pathway to capture  $\text{CO}_2$  and CO to make ethanol as well as acetic acid.
- A New Zealand start-up company **LanzaTech** developed a process with a strain that makes mainly ethanol using the off-gases from steel mills.
- It uses a specially designed fermentation vessel and several working-scale versions have been installed in steel mills and the like.
- The ethanol can be converted to various chemical precursors, and there is also a chemical process to convert it to ‘sustainable’ aviation fuel.

## ABOUT

### Carbon Recycling Technology for Today and the Future

LanzaTech's carbon recycling technology is like retrofitting a brewery onto an emission source like a steel mill or a landfill site, but instead of using sugars and yeast to make beer, pollution is converted by bacteria to fuels and chemicals! Imagine a day when your plane is powered by recycled GHG emissions, when your shampoo bottle started life as emissions from a steel mill. This future is possible today using LanzaTech technology.



Startup Year

**2022**

Annual Ethanol Production Volume

**60,000 Metric Tons**

Annual CO<sub>2</sub> Abatement Volume

**~120,000 Metric Tons**

Carbon Source

**Ferroalloy Mill Emissions**



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## Channelling Metabolism into Desired Routes

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- Ethanol from Plant Waste
- A Demonstrated Approach
- The Metabolic Model
- The Targets
- The Targets on the Map
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# Channelling Metabolism into Desired Routes

# Ethanol from Plant Waste

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#### ● Ethanol from Plant Waste

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Some of the issues:

- Plant wastes (e.g. straw) contain cellulose and hemicellulose which can be hydrolysed to glucose and pentose sugars.
- Yeasts convert glucose to ethanol, but don't readily use the pentoses.
- *Escherichia coli* can use pentoses as well as glucose, but ethanol is not its preferred product.
- *E. coli* is easy to engineer, but can it be modified to make ethanol from pentoses in such a way that it cannot mutate back to its original state?

# A Demonstrated Approach

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- Friedrich Sreenc's group (Trinh et al, Appl. Env. Microbiol, 74, 3634-3643, 2008) built a medium-sized structural model of E coli central carbon metabolism.

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- They computed the elementary modes leading from glucose and pentoses to products including ethanol and biomass.



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- They searched for reactions that were *needed* for the most number of modes leading to other products but which *still kept some* of the routes to biomass and ethanol.

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- They searched for reactions that were *needed* for the most number of modes leading to other products but which *still kept some* of the routes to biomass and ethanol.
- They found a set of *seven* reactions (eight genes) that between them disabled all the modes except those leading to either ethanol alone or biomass and ethanol.

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- They found a set of *seven* reactions (eight genes) that between them disabled all the modes except those leading to either ethanol alone or biomass and ethanol.
- They successively made the set of deletions where growth can only occur with ethanol production and obtained close to the theoretically-predicted yields of ethanol on glucose and xylose.

# The Metabolic Model

## Outline

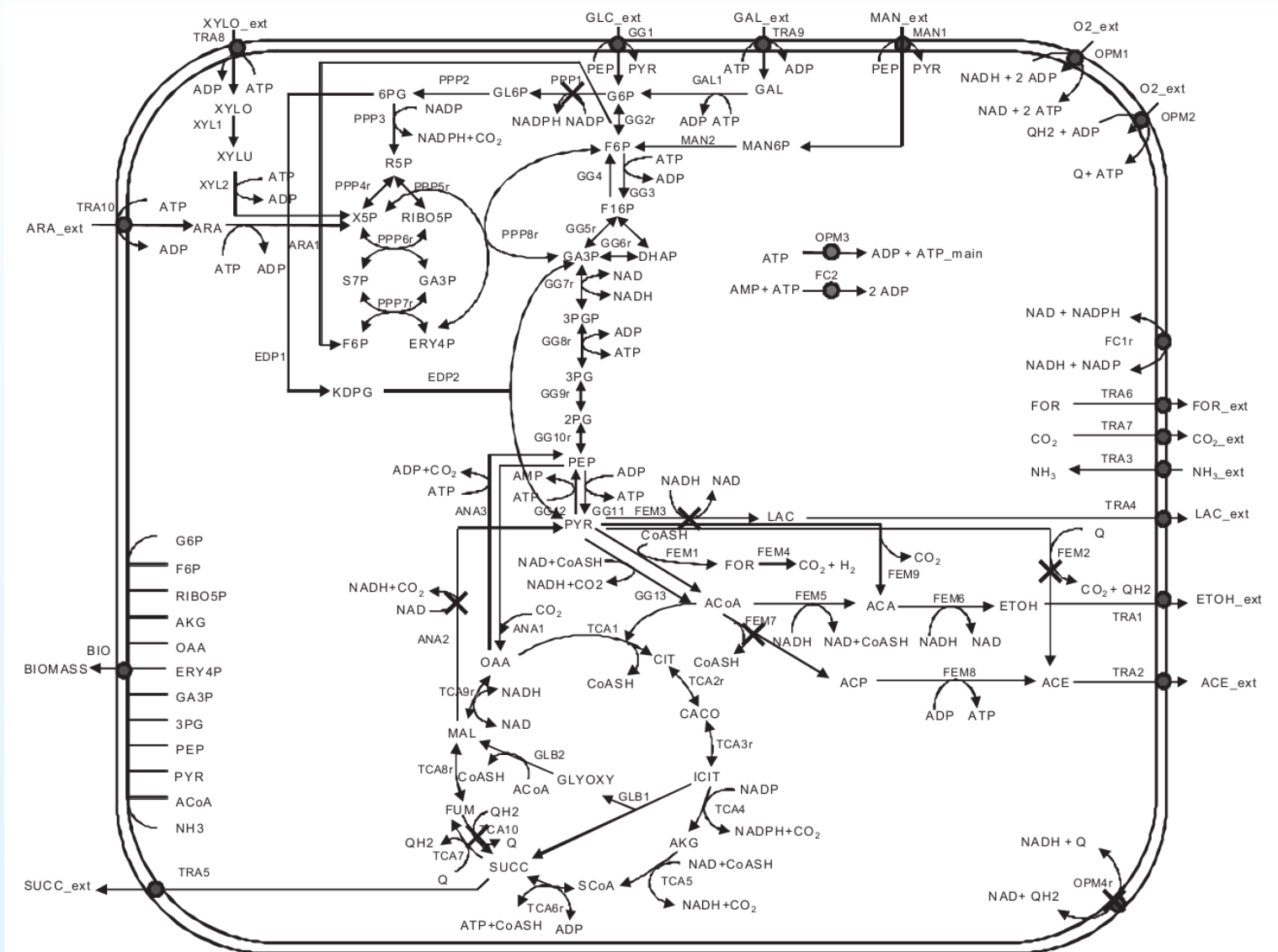
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# The Targets

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## Enzyme reaction

## Gene(s)

Glucose-6-P DH

*zwf*

NADH DH II

*ndh*

NAD/NADP malic enzyme

*sfcA, maeB*

D-lactate DH

*ldhA*

fumarate reductase

*frdA*

pyruvate oxidase

*poxB*

Pi acetyl transferase

*pta*

# The Targets on the Map

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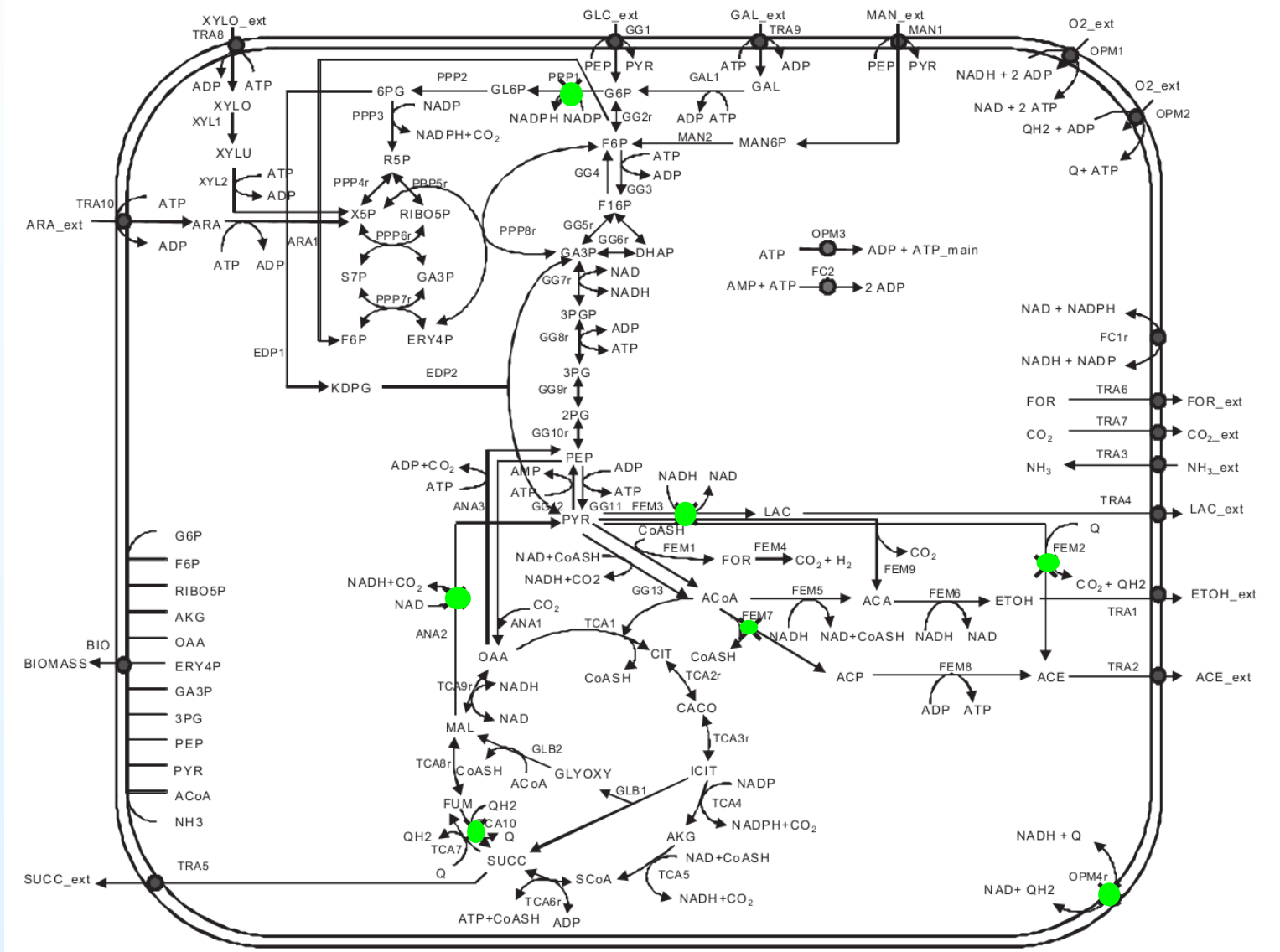
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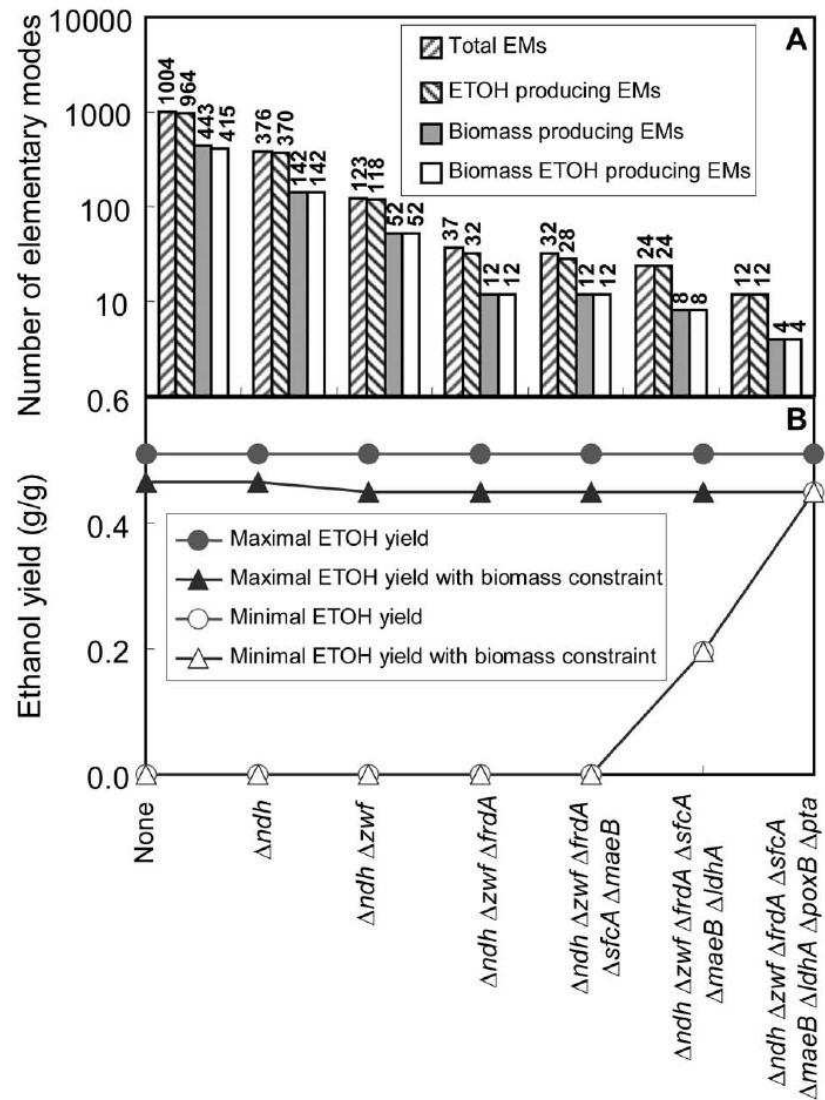
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- **Conclusion**
- Acknowledgements

- Elementary modes analysis can assist biotechnology projects to design metabolic network modifications for new products or to obtain improved yields.
- Strategies can include both addition of heterologous enzymes to provide new routes, or deletion of native enzymes to block unproductive routes.



## Acknowledgements

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